EDITORIAL



A triple sense of oxygen promotes neurovascular angiogenesis in NG2-derived cells

Expansion and remodelling of the cerebral vasculature via the induction of pro-angiogenic genes is a crucial issue in many pathologies. Urrutia et al¹ demonstrate how NG2-derived cells in the brain sense oxygen to promote neurovas-cular angiogenesis.

Master regulator of cellular adaptation to hypoxia, which includes angiogenesis, is the Hypoxia inducible factor (HIF). Three isoforms exist of the cellular oxygen sensor: HIF prolyl hydroxylases (PHD)-1,-2 and -3 (EGLN-2, -1, -3, respectively), enzymes that hydroxylate prolyl residues of the regulatory subunit HIF- α ,²⁻⁴ which, thus tagged, is easily captured by the von Hippel Lindau protein (VHL),⁵ and subsequently degraded by the proteasome (review in Ref.6). PHDs are ubiquitously expressed, as is the HIF system. Highly specific, synthetic, orally available, small molecule PHD-inhibitors are being tested in large phase III clinical trials for treatment of renal anaemia (review in Ref.7). Historically, erythropoietin (EPO) led to the discovery of HIF⁸ and PHD,^{2,3} while in the future, PHD-inhibitors may complement or even replace EPO. PHD inhibitors are potent erythropoiesis stimulating agents, comparable to EPO and its related compounds.9 Moreover, preclinical data suggest that PHD inhibition can improve tissue responses to hypoxia, and this may ameliorate highly relevant clinical conditions like ischemic stroke (review in Ref.10), myocardial infarction (review in Ref.11), acute kidney injury (review in Ref.12) etc Major outcome data from the above mentioned clinical trials are pending. In the meantime, it's worth taking a closer look at PHD inhibition in basic science, as it likely will help us understand upcoming clinical data. In this regard, Urrutia et al¹ present a meticulous assessment of brain angiogenesis, a highly relevant topic in ischemic and degenerative central nervous disorders.¹³ With help of knockout technology ranging from single to quadruple, they dissect the contributions of PHD-1 to -3 and EPO in this process. To the best of our knowledge, this is the most thoroughly conducted study addressing this topic. But, before we review their results, we need a little closer look at the HIF-PHD system: The transcription factor HIF is a heterodimer of an oxygen-dependent, variable alpha subunit (HIF-1 α , -2 α or -3 α) and a constitutive beta subunit (HIF- β , ARNT). HIF-1 α and -2 α , the most important and best studied alpha subunits, each have two oxygen dependent degradation domains (ODD), one at the C-terminus (CODD), and another at the N-terminus (NODD) (review in Ref.6). PHDs are dioxygenases using molecular O₂ as substrate, and 2-oxoglutarate as co-substrate. Their KmO₂ is above normal arterial O₂ concentration.¹⁴ PHD-2 is considered the most important isoform, responsible for HIF-α degradation in normoxia. PHD-3 and -2 are HIF target genes, and hence, as a negative feedback loop, help terminate the HIF response engendered by hypoxia. The four clinically tested PHD inhibitors roxadustat, vadadustat, daprodustat and molidustat are highly specific, but differ in kinetics and their ability to target HIF-1 α vs -2 α , and the respective CODD vs NODD.¹⁵ Hence, at least to a certain extent, the four compounds may exhibit different clinical effects. As a transcription factor, HIF acts through its target genes, the precise number of which remains unclear. Confusingly, the consensus DNA binding motif for HIF, the so-called hypoxia response element (HRE; RCGTG, with R = A or R = G) appears more than one million times in the genome.¹⁶ Clearly, most of these potential HREs are not accessible to HIF, but covered by chromatin. To make things more complex, some HREs of proven relevance locate quite far from the transcription initiation site. Furthermore, for full gene transactivation, binding of HIF to a single HRE seems insufficient, but rather requires binding to multiple HREs, or contribution from additional co-factors. Chromatin binding assays suggest that both HIF-1 α and HIF-2 α regulate some hundreds of genes, with partial overlap.¹⁷ Hypoxia response, thus, varies between underlying conditions, tissues and even individual cells. Altogether, interventions into the HIF-PHD system, whether pharmacological or genetical, are nearly unpredictable, which brings us back to the in vivo studies of Urrutia et al¹: They chose NG2-Cre mice to specifically knockout genes of interest in central nervous cells with particular relevance to angiogenesis, namely pericytes and NG2 glia cells. VHL knockout, a robust model for combined HIF-1 α and HIF-2 α up-regulation, led to enhanced angiogenesis and EPO. In the past, the latter has been attributed angiogenic effects in the brain, but a plethora of clinical trials (review in Ref.18) proved that exogenous EPO was unable to ameliorate brain damage. By double knockout (VHL/EPO)

^{© 2020} Scandinavian Physiological Society. Published by John Wiley & Sons Ltd

cta Physiologica

Urrutia et al¹ show that the angiogenic phenotype is independent of EPO. Interestingly, knockout of PHD-2, which carries the workload of HIF degradation in normoxia, did not lead to angiogenesis while double knockout (PHD-2/ PHD-3) did. The authors speculate that PHD-3, as a HIF target gene, had compensated for the single PHD-2 knockout. Knockout of PHD-1 alone had no angiogenic effect, most likely since preserved PHD-2 activity was sufficient to degrade HIF. However, combined knockout of all three PHD isoforms further enhanced angiogenesis with respect to the PHD-2/PHD-3 knockout. Hence, as the authors put it, each PHD isoform has its contribution to neurovascular homeostasis. And, finally, quadruple knockouts (PHD-1/PHD-2/ PHD-3/HIF-2 α) proved that the angiogenic phenotype was fully dependent on HIF-2 α .

Aside from sophisticated and refined methodology, the study by Urrutia et al¹ broadens our vision of the HIF-PHD components working together to shape a potentially relevant clinical effect. It seems that, indeed, cells make use of their triple oxygen sensor.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

Christian Rosenberger¹ D Michael Fähling²

¹Department of Nephrology and Renal Transplantation, Charité – Universitätsmedizin Berlin, corporate member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany

²Department of Vegetative Physiology, Charité – Universitätsmedizin Berlin, corporate member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany

Correspondence

Michael Fähling, Department of Vegetative Physiology, Charité – Universitätsmedizin Berlin, corporate member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany. Email: michael.faehling@charite.de

ORCID

Christian Rosenberger D https://orcid. org/0000-0003-4194-561X Michael Fähling D https://orcid.org/0000-0003-1079-5049

REFERENCES

- Urrutia AA, Guan N, Mesa-Ciller C, Afzal A, Davidoff O, Haase VH. Inactivation of HIF-prolyl 4-hydroxylases 1, 2 and 3 in NG2expressing cells induces HIF2-mediated neurovascular expansion independent of erythropoietin. *Acta Physiol (Oxf)*. 2020:e13547. https://doi.org/10.1111/apha.13547
- Ivan M, Kondo K, Yang H, et al. HIFalpha targeted for VHLmediated destruction by proline hydroxylation: implications for O2 sensing. *Science*. 2001;292:464-468.
- Jaakkola P, Mole DR, Tian YM, et al. Targeting of HIF-alpha to the von Hippel-Lindau ubiquitylation complex by O2-regulated prolyl hydroxylation. *Science*. 2001;292:468-472.
- Taylor MS. Characterization and comparative analysis of the EGLN gene family. *Gene*. 2001;275:125-132.
- Maxwell PH, Wiesener MS, Chang GW, et al. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygendependent proteolysis. *Nature*. 1999;399:271-275.
- Kling L, Schreiber A, Eckardt KU, Kettritz R. Hypoxia-inducible factors not only regulate but also are myeloid-cell treatment targets. *J Leukoc Biol.* 2020. https://doi.org/10.1002/JLB.4RI0820-535R
- Sanghani NS, Haase VH. Hypoxia-inducible factor activators in renal anemia: current clinical experience. *Adv Chronic Kidney Dis*. 2019;26:253-266.
- Wang GL, Semenza GL. Purification and characterization of hypoxia-inducible factor 1. J Biol Chem. 1995;270:1230-1237.
- Chen N, Hao C, Liu BC, et al. Roxadustat treatment for anemia in patients undergoing long-term dialysis. N Engl J Med. 2019;381:1011-1022.
- 10. Leu T, Schutzhold V, Fandrey J, Ferenz KB. When the brain yearns for oxygen. *Neurosignals*. 2019;27:50-61.
- Schreiber T, Salhofer L, Quinting T, Fandrey J. Things get broken: the hypoxia-inducible factor prolyl hydroxylases in ischemic heart disease. *Basic Res Cardiol.* 2019;114:16.
- Shu S, Wang Y, Zheng M, Liu Z, Cai J, Tang C, Dong Z. Hypoxia and hypoxia-inducible factors in kidney injury and repair. *Cells*. 2019;8:207.
- Hatakeyama M, Ninomiya I, Kanazawa M. Angiogenesis and neuronal remodeling after ischemic stroke. *Neural Regen Res.* 2020;15:16-19.
- Koivunen P, Hirsila M, Gunzler V, Kivirikko KI, Myllyharju J. Catalytic properties of the asparaginyl hydroxylase (FIH) in the oxygen sensing pathway are distinct from those of its prolyl 4-hydroxylases. *J Biol Chem*. 2004;279:9899-9904.
- Yeh TL, Leissing TM, Abboud MI, et al. Molecular and cellular mechanisms of HIF prolyl hydroxylase inhibitors in clinical trials. *Chem Sci.* 2017;8:7651-7668.
- Wenger RH, Stiehl DP, Camenisch G. Integration of oxygen signaling at the consensus HRE. *Sci STKE*. 2005;2005;re12.
- Schodel J, Mole DR, Ratcliffe PJ. Pan-genomic binding of hypoxia-inducible transcription factors. *Biol Chem.* 2013;394: 507-517.
- Siren AL, Fasshauer T, Bartels C, Ehrenreich H. Therapeutic potential of erythropoietin and its structural or functional variants in the nervous system. *Neurotherapeutics*. 2009;6:108-127.